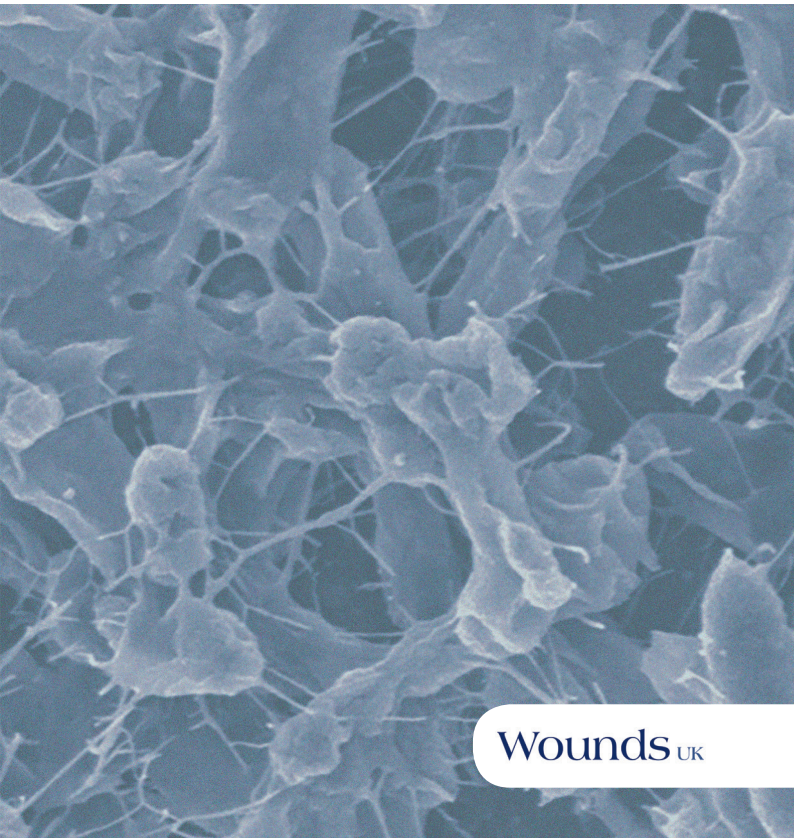




QUICK GUIDE

MANAGING BIOFILM IN STATIC WOUNDS



Wounds^{UK}

UNDERSTANDING BIOFILM

Biofilm has been found to be present in a majority of static wounds¹ and may be a key cause of delayed wound healing² and a precursor to infection.³

Biofilms are complex microbial communities containing micro-organisms, embedded in a protective, slimy barrier of sugars and proteins.

Biofilm can protect micro-organisms from the host immune response and from antimicrobial agents, protecting micro-organisms and allowing them to multiply. In addition, biofilm is difficult to completely remove,⁴ even with debridement, and it can reform quickly.⁵

Because of the variability and complexity of biofilm structure, visual observation of wound bioburden can be challenging. Specialist diagnostic testing is not readily available.⁶

A 'shiny' or 'slimy' wound surface, persistence of slough-like material and stalled healing may indicate the presence of biofilm. Early identification and management of biofilm in a wound can improve wound healing and patient wellbeing.⁶

Managing wounds containing biofilm:⁷

Adopt strategies to reduce the amount of biofilm and help prevent its reformation.

Address factors that may contribute to wound chronicity, such as wound infection and moisture imbalance.

Follow a protocol of care that incorporates cleansing and/or debridement, and select an appropriate antimicrobial dressing.

1. James GA et al (2008) Biofilms in chronic wounds. *Wound Rep Regen* 16:37-44
2. Metcalf D, Bowler P (2013) Biofilm delays wound healing: A review of the evidence. *Burns & Trauma* 1:5-12
3. Percival SL, Bowler PG (2004) Biofilms and their potential role in wound healing. *Wounds* 16:234-40
4. Wolcott RD et al (2010) Biofilm maturity studies indicate sharp debridement opens a time-dependent therapeutic window. *J Wound Care* 19:320-8
5. Wolcott RD et al (2009) Regular debridement is the main tool for maintaining a healthy wound bed in most chronic wounds. *J Wound Care* 18:54-6

6. Phillips et al (2010) Biofilms Made Easy. *Wounds Int*
7. Parsons D et al (2014) Next-generation antimicrobial dressings: AQUACEL® Ag+ Extra and Ribbon. *Wounds Int*
8. Metcalf D et al (2014) A clinical algorithm for wound biofilm identification. *J Wound Care* 23(3):137-43
9. Physical Disruption of Biofilm by AQUACEL® Ag+ Wound Dressing Scientific Report. WHR13850 MA232. 2013. Data on File, ConvaTec Inc.
10. Antimicrobial activity and prevention of biofilm reformation by AQUACEL™ Ag+ EXTRA dressing. Scientific Background Report. WHR13857, MA236, 2013. Data on file, ConvaTec Inc.

AQUACEL® Ag+ dressings

Demonstrated wound healing in a real-life evaluation of clinical cases¹¹

113

113 cases of challenging, at-risk or infected wounds

74%

74% had suspected biofilm

95%

95% of wounds improved or healed

4.1

4.1 weeks average management period

Ag+ Technology

Ag+ Technology is a unique, silver-containing formulation¹² that:

- disrupts and breaks down biofilm slime to expose bacteria^{*9,10,13}
- kills a broad spectrum of bacteria, including antibiotic resistant superbugs, with its reservoir of silver^{†*10,13,14}
- prevents biofilm reformation^{*10,13}

Hydrofiber® Technology

Helps create an ideal environment for healing, and for the Ag+ Technology to work

- Locks in excess exudate and bacteria to help minimise cross-infection and prevent maceration^{*15-18,19,20}
- Micro-contours to the wound bed, helping to maintain optimal moisture balance and eliminating dead spaces where bacteria and biofilm can develop^{*21-23}
- Responds to wound conditions by forming a cohesive gel, while helping minimise pain associated with dressing changes^{*24-26}

Supported by ConvaTec | www.convatec.co.uk

*As demonstrated in vitro / †including MRSA, VRE and ESBL bacteria

17. Bowler PG et al (1999) Infection control properties of some wound dressings. *J Wound Care* 8:499-502
18. Walker M et al (2007) In vitro studies to show sequestration matrix metalloproteinases by silver containing wound care products. *Ostomy Wound Manage* 53:18-25
19. Walker M, Parsons D (2010) Hydrofiber technology: its role in exudate management. *Wounds UK* 6(2):31-8
20. Parsons D et al (2005) Silver antimicrobial dressings in wound management: A comparison of antibacterial, physical and chemical characteristics. *Wounds* 17:222-32
21. Jones SA et al (2005) Antimicrobial activity of silver-containing dressings is influenced by dressing conformability with a wound surfact. *Wounds* 17:263-70
22. Bowler P et al (2010) Dressing conformability and silver-containing wound dressings. *Wounds UK* 6(2):14-20
23. Walker M et al (2011) Evaluation of low-adherent antimicrobial dressings. *Wounds UK* 7(2):32-45
24. Barnea Y et al (2004) Clinical comparative study of Aquacel and paraffin gauze dressing for split-skin donor site treatment. *Ann Plast Surg* 53:132-6
25. Kogan L et al (2004) Comparative study of Aquacel and Silverol treatment in burns. *Ann Burns Fire Dis* 17:201-7
26. Brunner U et al (2000) Experiences with hydrofibres in the moist treatment of chronic wounds, in particular of diabetic foot. *VASA* 29:253-7

CLINICAL ALGORITHM FOR BIOFILM IDENTIFICATION⁸

Routine assessment of static wounds should include a thorough review that incorporates visual and indirect indicators to identify suspected biofilm and guide management. This algorithm (developed by ConvaTec Ltd), helps identify biofilm.⁸

Wound: visual indicators

1. Does the surface material detach easily and atraumatically from the underlying wound bed using physical removal techniques such as swabs, pads or sharp debridement?

Yes

No

Probably biofilm
with increasing
confidence

Probably host
devitalised
tissue (e.g.
slough, fibrin)

2. Does the surface material persist and/or reform quickly (in 1-2 days) despite intervention (e.g. debridement, cleansing)?

Yes

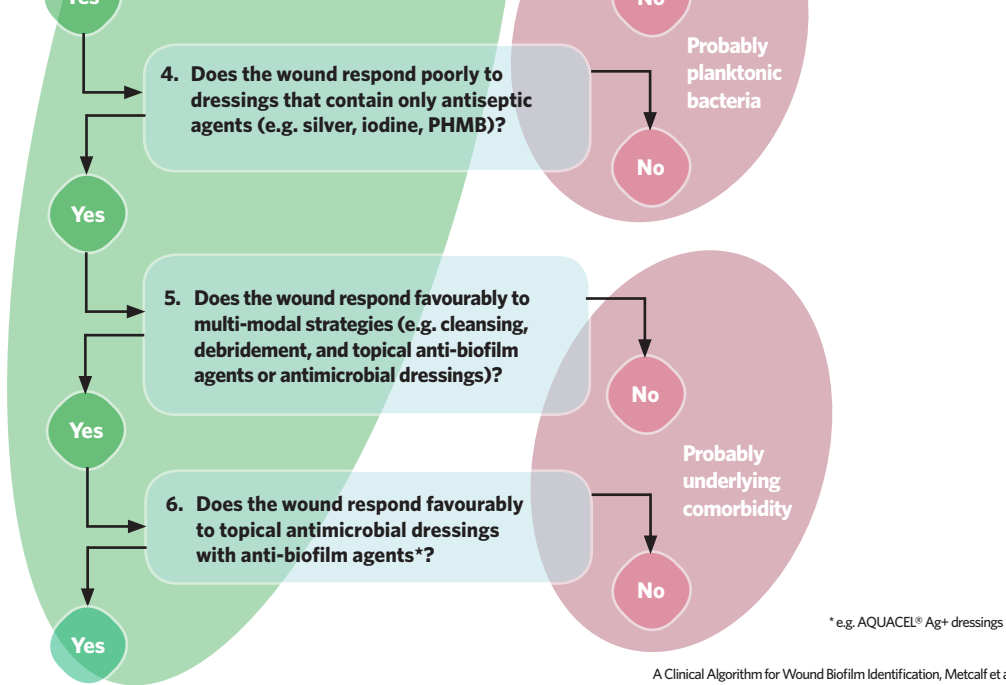
No

Wound: indirect indicators

3. Does the wound respond poorly to topical or systemic antibiotics?

Yes

No



A Clinical Algorithm for Wound Biofilm Identification, Metcalf et al.
Adapted from Journal of Wound Care 2014; 23(3): 137-143

11. Walker et al (2015) A real-life clinical evaluation of a next-generation antimicrobial dressing on acute and chronic wounds. J Wound Care 24:1, 11-22

12. Composition comprising antimicrobial metal ions and a quarternary cationic surfactant. Scientific Background Report. WO 2012136968 A1, 2012. Data on file, ConvaTec Inc.

3-STEP PROTOCOL OF CARE⁷

ASSESS

Evaluate both the patient and the wound

- Carry out a holistic patient assessment (e.g. medication, comorbidities, lifestyle issues)
- Assess the wound:
 - Wound type and length of time wound has been present
 - Wound bed appearance (tissue type and percentage of: slough, necrosis, granulation, suspected biofilm)
 - Size (length, width, depth)
 - Exudate (colour, consistency, level)
 - Associated pain and/or odour
 - Peri-wound skin condition (swelling, discolouration, maceration)
 - Signs/symptoms of infection (pain, odour, heat, redness, swelling, purulence)

MANAGE

Cleanse and debride

- Cleanse and debride the wound where necessary to remove barriers to healing (e.g. slough, necrosis, biofilm)
- Dress the wound:
 - Apply an appropriate dressing that can disrupt biofilm, kill bacteria and prevent biofilm reformation, while managing exudate and infection (e.g. AQUACEL[®] Ag+ dressings)⁷

MONITOR

Reassess and document the wound at each dressing change

- If the wound remains infected or at risk of infection, continue to use a suitable dressing such as AQUACEL[®] Ag+ Extra[™] dressing or AQUACEL[®] Ag+ Ribbon dressing covered with a secondary dressing such as AQUACEL[®] Foam dressing

*As demonstrated in vitro

13. Antimicrobial activity against CA-MRSA and prevention of biofilm reformation by AQUACEL[™] Ag+ EXTRA[™]. Scientific Background Report. WHR13875 MA239, 2013. Data on file, ConvaTec Inc.
14. Bowler PG et al (2012) Multi-drug resistant organisms, wounds and topical anti-microbial protection. Int Wound J 9:387-96

15. Newman GR et al (2006) Visualisation of bacterial sequestration and bacterial activity within hydrating Hydrofiber[™] wound dressings. Biomaterials 27:1129-39
16. Walker M et al (2003) Scanning electron microscopic examination of bacterial immobilization in a carboxymethyl-cellulose (AQUACEL[™]) and alginate dressing. Biomaterials 24:883-90