

Technology update: role of wound cleansing in the management of wounds

KEY WORDS

- ▶ Biofilm
- ▶ Cleansing
- ▶ Irrigation

Wound cleansing should be seen as an integral part of wound bed preparation to optimise the wound environment by removing debris, reducing bacterial load and preventing biofilm activity. Clinicians have a number of options to choose from when selecting an appropriate wound irrigation solution. One factor to consider includes the ability to mount a rapid antimicrobial response to microbial contamination, while avoiding damage to human cells important for wound healing.

Micro-organisms produce significant barriers to the healing of chronic wounds (Kim et al, 2010). They are present in all wounds, although not all wounds become chronic. Microbes grow in both a planktonic phenotype (free-floating mobile single cells) and biofilm phenotype (a fixed polymicrobial community; James et al, 2008). A self-secreted matrix protects the biofilm from attack by the host immune system and makes the wound harder to treat (Tuttle et al, 2011). In fact, many of the characteristics of chronic wounds, such as persistent inflammation, exudate, and host-cell senescence, result directly from biofilm (Katsuyama et al, 2005; Wolcott et al, 2008; Raymond et al, 2013).

Wounds that have become chronic — characterised by delayed or stalled healing — due to suspected biofilm/increased bioburden should be managed with a biofilm-based approach to wound care (Wolcott et al, 2010). This means combining wound cleansing/irrigation with debridement and application of topical antimicrobial agents (e.g. wound cleansers and dressings), to facilitate healing by disrupting, removing and preventing the reformation of biofilm (Rosdahl and Kowalski, 2008).

IMPORTANCE OF WOUND BED PREPARATION

Wound bed preparation is recognised as having a key role in wound management. The International Advisory Board on Wound Bed Preparation has developed an assessment tool, known by the acronym TIME (T = tissue, non-viable or deficient; I = infection or inflammation; M =

moisture imbalance; E = edge of wound, non-advancing or undermined), that sets out the goals of wound-bed preparation: removing non-viable tissue, reducing oedema and exudate, reducing the bacterial burden and correcting any abnormalities to promote wound healing (Falanga, 2004). Wound bed preparation offers a structured and systematic approach to assist clinicians when assessing and managing patients with wounds.

WHERE WOUND CLEANSING FITS

Wound cleansing (*Box 1*) can help achieve the goals of wound bed preparation by assisting in removing loose material to create the optimal local conditions for wound healing by removing exudate and other debris. Irrigation is the preferred method of wound cleansing, as it can clear the wound of debris and microbes, while avoiding trauma in the wound bed (Gabriel et al, 2013).

A practical strategy for wound bed preparation in chronic wounds is to lightly irrigate the wound before inspection and assessment. Debride any soft, degraded areas of the wound bed, slough or necrotic tissue according to local protocols, and then re-irrigate the wound with a cleansing solution before applying an appropriate dressing, also according to local protocols.

There are several broad categories of solutions that can be used: saline and water, highly reactive solutions, and minimally or non-cytotoxic antimicrobial-containing solutions.

EXPLORING IRRIGATION SOLUTIONS

When considering the clinical benefits of wound

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irrigation and the appropriate agents to use, the clinician should keep in mind that an ideal wound irrigation solution will provide periodic reduction of bacterial contamination and removal of debris without adversely affecting cellular activities crucial to the wound-healing process (Hoon et al, 2013). It is, therefore, important to consider the balance of antimicrobial action and cytotoxicity when choosing a wound irrigation solution (Table 1).

SALINE AND TAP WATER

Although tap water and saline are not cytotoxic and do not seem to be harmful to wounds (Fernandez et al, 2002), these cleansing choices may not actively promote healing, particularly in chronic wounds with biofilm and/or increased bacterial burden (Cutting, 2010), indicating that non-antiseptic cleansing does not remove harmful molecules such as matrix metalloproteases and elevated proinflammatory cytokines. These molecules may be present in chronic wounds and delay healing.

Although these agents have been used as standards of non-cytotoxicity (Wilson et al, 2005), neither has significant antimicrobial properties. This means saline is not appropriate for augmenting wound-bed preparation in the context of biofilm-based wound care (Leaper et al, 2012), or where wounds are clinically diagnosed as locally infected or at risk of infection. Further, it should be pointed out that even ‘safe’ tap water can become colonised with viable microbes. In particular, *Pseudomonas* is well-documented in the plumbing systems of healthcare facilities (Trautmann et al, 2005; Mena and Gerba, 2009; Jefferies et al, 2012). Therefore, a cleansing agent with broad-spectrum activity and rapid kill rates (dwell time of 5 to 10 minutes) should be chosen to aid wound bed preparation.

HIGHLY REACTIVE SOLUTIONS

Solutions such as peroxide and povidone iodine (depending on the carrier and the strength of

the povidone iodine) and commercially available products (e.g. alcohol-based cleansers, soaps, foams and wipes) can be particularly cytotoxic (Sibbald et al, 2011). A 2010 Cochrane review suggested that such solutions may do little to control wound bacteria and may in fact interfere with host healing mechanisms (Fernandez et al, 2002). Although they probably do not significantly harm the host, the high level of reactivity is not necessary.

ANTIMICROBIAL SOLUTIONS

Multiple antiseptic agents are available and have been extensively evaluated for cytotoxicity and their biocidal abilities against a broad spectrum of microorganisms, including bacteria and fungus (yeast; Leaper et al, 2012). Common cleansing agents such as octenidine and polyhexamethylene biguanide (PHMB) are minimally toxic to host cells when used in low concentrations (Rani et al, 2014).

Hypochlorous acid is produced by the body’s immune cells in response to invading pathogens (Eryilmaz and Palabruyik, 2013). It is available as a commercially-prepared wound cleansing solution, which is non-toxic, effective against a broad range of microorganisms and has a rapid kill rate (Wang et al, 2007; Crew et al, 2012). Clinical studies of hypochlorous acid demonstrate improved wound-healing outcomes (Robson et al, 2007; Crew et al, 2012).

RATIONALE FOR ANTIMICROBIAL SOLUTIONS

After a wound has been washed and debrided, exposing microbial cells to the environment, the antimicrobial properties of a cleansing agent can be effective. Because microbes may exist on the wound bed in high numbers, agents stronger than simple saline or tap water may be needed. Antimicrobial solutions can be used as part of the balance in managing wound colonisation, biofilm and/or infection. Non-toxic agents that are effective at low concentrations should be considered when possible.

Antimicrobial solutions have the added benefit of being usable across the spectrum of chronic wounds without resulting in bacterial resistance issues, because they are antiseptic rather than antibiotic. These factors, along with the ability to disrupt and prevent reformation of biofilm/reduce bioburden are ideal properties of an antimicrobial

Box 1. Definition of wound cleansing

Remove surface contaminants, loose debris, slough, softened necrosis, microbes and/or remnants of previous dressings from the wound surface and its surrounding skin (Rodeheaver and Ratliff, 2007).

irrigation solution (Figure 1). As a result, such irrigation solutions may have an important place within the process of chronic wound management.

SUMMARY

Resolving critical colonisation, biofilm and/or infection in a chronic wound is achieved through appropriate wound bed preparation. The goal of wound cleansing is to remove loose material in the wound bed, reduce bacterial load and to assist in the suppression of biofilm.

Cleansing should be tailored according to the goals determined by holistic assessment of the patient and wound, and best practices for irrigation followed (Box 2). Generally speaking, wounds

should be irrigated at every dressing change until visible debris is removed. Cleansing agents may also be useful in removing encrusted dressings in order to lift them and avoid causing trauma to newly formed granulation tissue in the wound bed.

It is critical that the wound cleansing agent does not impair the wound healing process, but is strong enough to remove the protective matrix secreted by microbes attached to the wound (biofilm) and reduce the bioburden.

Clinicians have a number of options when selecting a wound cleansing agent (Figure 1). Although saline and water have been found by numerous studies to not be harmful to wounds, this feature is not enough in the context of biofilm-based wound care. Other

Table 1. Key considerations in some common wound irrigation agents.

Saline (Gabriel et al, 2013)	Low toxicity Limited ability to reduce bacterial load Bacterial growth can occur in an open container within 24 hours
Sterile water (Gabriel et al, 2013)	Limited ability to reduce bacterial load Readily absorbed by tissues; water toxicity may result when excess volumes are used No longer sterile after opening
Tap water (Mena et al, 2009; Gabriel et al, 2013)	Recommended where saline and sterile water are not available Limited ability to reduce bacterial load Microbes, in particular <i>P. aeruginosa</i> , can colonise taps and as a result may end up in wounds irrigated in this way
Commercially available products (e.g. foams, soaps, wipes and solutions with surfactants; Gabriel et al, 2013)	Remove bacteria with less required force due to surfactant content May be best suited for wounds with adherent cellular debris and biofilm Typically contain preservatives to extend effective shelf life Can be highly cytotoxic to healthy cells and granulating tissue
Povidone iodine (Gabriel et al, 2013)	Broad-spectrum antimicrobial activity Cytotoxic to healthy cells and granulating tissue in higher-percentage concentrations May irritate periwound skin
Hydrogen peroxide (Gabriel et al, 2013)	May be cytotoxic to healthy cells and granulating tissue Ineffective in reducing bacterial counts
Polyhexamethylene biguanide (PHMB) 0.1% (Bradbury and Fletcher, 2011)	Also contains betaine, a surfactant, to lift microbes and debris and suspend them in solution to prevent wound recontamination Has an increased ability to penetrate difficult-to-remove coatings, lifting debris, bacteria and biofilm from the wound Broad spectrum of activity against bacteria, viruses and fungi No evidence of resistance
Octenidine (Braun et al, 2013)	Can be used to loosen encrusted dressings in addition to irrigating debris and microbes from the wound bed Contains octenidine dihydrochloride, a preservative, to extend shelf life and a surfactant-like molecule that is effective at infiltrating wounds while being less irritating Shown to prevent and remove the growth of bacterial biofilms
Hypochlorous acid 0.01% (Shetty et al, 1999; Wang et al, 1999; Zinkevich et al, 2000; Selkon et al, 2006; Niezgoda et al, 2010; Hoon et al, 2013; Martindale Pharma, data on file)	Broad-spectrum antimicrobial activity Non-irritating, non-sensitising, non-toxic Can be used to loosen encrusted dressings in addition to irrigating loose debris and bacteria from the wound bed Has rapid antimicrobial activity at concentrations safe for human cells

wound cleansers, such as povidone iodine (in higher concentrations), soaps, peroxide and alcohol may be too harsh (Rabenberg et al, 2002).

The ideal wound irrigation solution should exhibit potent and rapid antimicrobial activity at concentrations that do not damage host cells required for wound healing. Hypochlorous acid, when used in low concentrations, is non-toxic and has a rapid antimicrobial action. The choice of

irrigation solution should also reflect the individual requirements of the wound and the patient after thorough holistic assessment (Cutting, 2010).

Where holistic assessment leads to the suspicion that bioburden is delaying wound healing, clinicians should consider using a suitable antimicrobial wound irrigation solution to promote healing, reduce bioburden, prevent the proliferation of biofilms and/or remove biofilm before applying

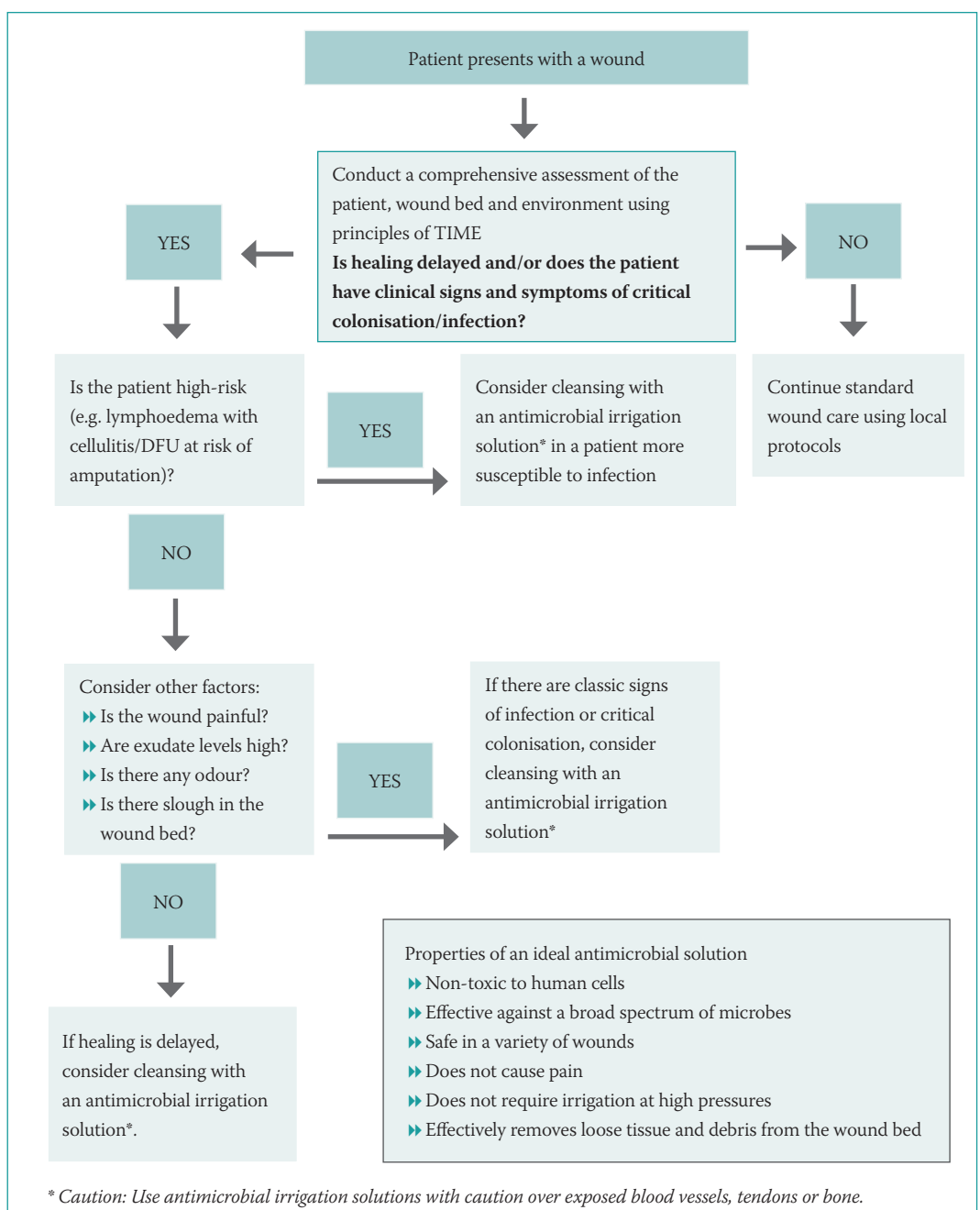


Figure 1. Wound-cleansing decision making pathway.

an appropriate dressing. As such, cleansing should become an integral part of wound-bed preparation, to help prevent infection and optimise the wound environment for healing. **WUK**

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Box 2. Practical tips for wound irrigation.

- ▶ Choice of solution: Base choice on assessment of both the patient (including medical condition and allergies) and the wound (e.g. clinically assessed as critically colonised; Gabriel et al, 2013; Sardina, 2013).
- ▶ Method of delivery: Deliver irrigation based on the needs of the patient (e.g. pain levels) and wound (e.g. fragility of wound and periwound skin; Gabriel et al, 2013; Sardina, 2013).
- ▶ Volume of solution: Volumes of 50–100 ml per centimetre of wound length is the general rule of thumb [35,36] (Chisholm et al, 1992; Lammers et al, 2003).
- ▶ Prevention of cross-contamination: The clinician should wear personal protective equipment. Do not use solution that has been opened for longer than 24 hours (Gabriel et al, 2013; Sardina, 2013).
- ▶ Comfort of patient: Make sure irrigation solution is at room temperature or slightly warmer. Use analgesia for painful wounds and allow time for it to take effect (Gabriel et al, 2013; Sardina, 2013).
- ▶ Irrigation of wound: Position the patient so the solution runs from the upper end of the wound downward or from clean to dirty (if the upper end is heavily infected and the lower end is clean), into a clean basin or irrigating pouch (Gabriel et al, 2013; Sardina, 2013).
- ▶ Documentation of treatment: Record all aspects of the wound cleansing, including assessment of the wound (e.g. slough, exudate, pain, erythema), date and time of treatment, amount and type of solution used, skin care performed, wound dressing(s) applied and notes on the patient's concordance with treatment (Gabriel et al, 2013; Sardina, 2013).

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